Straightforward syntheses of biradical-producing bicyclic dienediynes —dienediyne ketones cycloaromatize *via* the Saito–Myers and not *via* the neocarzinostatin pathway

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The dienediyne models 23 and 28 of the pharmacophore of the antitumor natural product neocarzinostatin were prepared. Each synthesis requires only six steps from α-formylcyclohexanone. Our approach uses two key steps. The first consists of one-pot biscoupling reactions between propargyl alcohol, 2,2-dimethyl-3-butyn-1-ol and the bis(enoltrifluoromethanesulfonate) 17. The second key step corresponds to ring-closing pinacol coupling reactions of the dialdehydes 20 and 25. The dienediyne models 23 and 28 cycloaromatized efficiently when treated with methyl thioglycolate and 1,4-cyclohexadiene at 25 °C via a Saito-Myers cyclization⁴ (to give the octahydroanthracenones 29, iso-29 and octahydrophenanthrenones 34, 35, respectively); in addition, we isolated compounds tentatively assigned as the octahydrobenzazulenones 30 and iso-30, which would stem from a competing Schmittel cyclization. According to density functional theory (B3LYP/6-31G*) and ab initio calculations [CASMP2(2.2)/6-31G//CAS(2.2)/6-31G], the core structures of the octahydroanthracenones and octahydrophenanthrenones obtained here and elsewhere form via the Saito-Myers cyclization of enyneallenyl ketones 53 to toluene-α, meta biradicals 55 and not via neocarzinostatin-like cycloaromatizations of the tautomeric enyne[3] cumulenols 54 to styrene \alpha, meta-biradicals 56. This is so because, on the one hand, the two cyclization modes are predicted to have similar activation barriers (Saito-Myers: 16.0 kcal mol⁻¹; neocarzinostatin type: 18.3 kcal mol⁻¹) but, on the other hand, the enyneallenyl ketone 53 is a much more stable (21.1 kcal mol⁻¹) cycloaromatization substrate than the enynecumulenol 54. In addition, the Saito-Myers cyclization product 55 is calculated to be considerably more stable (35.3 kcal mol⁻¹) than the neocarzinostatin-type cycloaromatization product 56.

The addition of neocarzinostatin, the calichemicins, the esperamicins, dynemicin A, kedarcidin, C-1027 and maduropeptin to the arsenal of anticancer agents has inspired many chemists to participate in a world-wide search for chemotherapeutics that function by similar mechanisms as these natural products. 1 Most of the latter compounds (the calichemicins, the esperamicins, dynemicin A, kedarcidin, C-1027) contain or form (maduropeptin) a highly strained 1,5hexadiyn-3-ene substructure, which isomerizes at body temperature or below to a benzene-1,4-biradical by a Bergman cyclization.² Neocarzinostatin is a highly strained dienediyne that can react to 7-octyne-1,2,3,5-tetraenes,3 which in turn cycloaromatize to styrene biradicals ('neocarzinostatin cyclization'). In 1989, Saito and Myers and their respective coworkers recognized that 6-heptyne-1,2,4-trienes (from now on 'enyneallenes', 3) can also cycloaromatize ('Saito-Myers cyclization', Scheme 1).4 They thereby form short-lived toluene-a, meta biradicals 4, which abstract hydrogen atoms from easily oxidizable compounds (1,4-cyclohexadiene, thiols or DNA) to give toluenes 6. Saito-Myers cyclizations have since then been encountered in many envneallenes 3,5 including envnallenyl ketones 3a,b.6-16 Recently, Schmittel et al.17 and Gillmann et al.18 found that some enyneallenes 3 undergo

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a different biradical-producing cyclization reaction, furnishing fulvene-1,3-diyls 5 ('Schmittel cyclization'). This increasingly observed 19 reaction mode is not a *cycloaromatization* and in this regard is distinct from Bergman, neocarzinostatin and Saito–Myers cyclizations. Whether a given enyneallene 3 cyclizes *via* the Saito–Myers (to give 4) or the Schmittel (to give 5) modes depends on whether the in-plane π orbitals of substrate 3 cause better 1,6 or 1,5 overlaps.

The common access (exception: ref. 16) to enynallenyl ketones 3a,b is the 1,6-addition of a nucleophile to dienediyne ketones 1 or the 1,8-addition of a nucleophile to dienediyne ketones 2 (Scheme 1). Previously synthesized type-1 dienediyne ketones were compounds 7-11,6-13 and type-2 dienediyne ketones were compounds 12–14^{14,15} (Scheme 2). The diene moieties of these dienediyne ketones are integrated into 5- (7-12) or 6-membered (13, 14) carbocycles. This ensures that derived enyneallenyl ketone 3 possess a cis C=C bond, which is a prerequisite for Saito-Myers cyclizations, $3 \rightarrow 4$. The diyne moieties of the dienediyne ketones 9-14 are part of 9- to 11-membered rings. This ensures that the in-plane π orbitals of derived enyneallenyl ketones 3 interact in a transannular fashion and thus particularly efficiently. Accordingly, the bicyclic dienediyne ketones in Scheme 2 cycloaromatize at lower temperatures and with higher yields (9: -30 °C/54%;8 **10a**: $20 \,^{\circ}\text{C}/31\%^{9a}$ or $25 \,^{\circ}\text{C}/55\%;^{9b}$ **10b**: $25 \,^{\circ}\text{C}/56\%;^{10}$ **10c**: $25 \,^{\circ}\text{C}/54\%;^{10}$ **10d**: $25 \,^{\circ}\text{C}/64\%;^{11}$ **10e**: room temp./28%;¹² **11**:

Scheme 2 Type-1 dienediyne ketones (upper rows) and type-2 dienediyne ketones (bottom row) from the literature

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 $0 \,{}^{\circ}\text{C}/20\%;^{13}$ **12**: $25 \,{}^{\circ}\text{C}/38\%;^{14}$ **13**: $35-37 \,{}^{\circ}\text{C}/24\%;^{15}$ **14**: $35-37 \,{}^{\circ}\text{C}/>35\%^{15}$) than the *monocylic* dienediyne ketone **7** $(70 \,{}^{\circ}\text{C}/15\%^6)$.

A dream of many researchers is that one day Saito-Myers cyclizations $3 \rightarrow 4$ will occur in synthetic anti-cancer agents endowed with tumor cell specificity by a yet unknown method. In this respect it would be desirable that biradical-forming bicyclic enyneallenyl ketones 3a,b be made available by paths that are as efficient and short as possible. In contrast to this, present-day synthesis path lengths for their dienediyne ketone precursors vary between 8 $(10a^9)$ and 15 steps $(10e^{12})$; the synthesis of the more elaborate and enantiopure neocarzinostatin model $10d^{11}$ required even 22 steps. We felt that there is not only a need but also leeway for improvement. We report below the presently shortest syntheses of dienediyne models of neocarzinostatin forming biradicals below body temperature. One leads to a type-1 and the other to a type-2 dienediyne ketone.

Results

The novel dienediyne ketone syntheses (Schemes 3, 4) use our bis(trifluoromethanesulfonate) ('bistriflate') \rightarrow dienediyne strategy. This strategy was originally developed for accessing neocarzinostatin models containing a cyclopentene ring. ²⁰ It was subsequently extended to the cyclohexene ring. ^{15,21,22} The starting material of these syntheses is the equilibrium mixture between the tautomers 15 and 16 of 2-formylcyclohexanone. Upon treatment with *tert*-butyllithium in THF the enol fraction forms a Z-configurated enolate. It is scavenged with triflic anhydride as a Z-monotriflate. ¹⁵ The lithium enolate formed from the latter compound and hexamethyldisilazide is sulfonylated with triflic anhydride once more. It thereby gives the stereopure bistriflate 17. ¹⁵

As already described²¹ and several times exploited, ^{15,22} bistriflate 17 couples under Pd catalysis with in situ formed copper acetylides via dienyne monotriflates to dienediynes. This occurs such that a fast first coupling reaction occurs at the C_{quat} —O bond. A much slower second coupling follows at the C_{tert} —O bond. Thus, successive additions of first 1.0 equiv of a first alkyne and a couple of hours later of ≥ 1.0 equiv of a second alkyne to bistriflate 17 lead to the formation of a dienediyne with predictable attachment sites of the alkynyl arms. Reversing the order in which the two alkynes are added is therefore tantamount to synthesizing a different dienediyne. Accordingly, coupling the bistriflate 17 in a one-pot reaction first with 2,2-dimethyl-3-butyn-1-ol and thereafter with propargyl alcohol gave the dienediynediol 19 (Scheme 3). The opposite order of couplings in the complementary one-pot reaction of Scheme 4 led to the dienediynediol 24. Unfortunately, the positional selectivities of the first couplings were not perfect. Thus, the desired bicyclic product 19 was contaminated with up to 10% of isomer 24 and the biscoupling product 24 with up to 10% of isomer 19. We coped with this imperfection best by purifying each biscoupling product several times by means of thorough flash chromatography on silica gel.²³ This is because compound 19 accumulates in the early and compound 24 in the late fractions. Alternatively, we isolated the monocoupling product 18 of bistriflate 17 and 2,2dimethyl-3-butyn-1-ol en route to the biscoupling product 19. However, separating 18 from 5-7% of the isomeric monocoupling product was difficult, too, and 18 polymerized more easily than directly formed 19.

The next step in the syntheses of Schemes 3 and 4 were oxidations of the dienediynediols **19** and **24** with the Dess-Martin reagent²⁴ to the dienediyne dialdehydes **20** [58% admixed with 10% of the *E* isomer; $\delta_{Z-C(=O)H} = 9.28$ (d, ${}^5J = 1.2$ Hz) and 9.52 (s)] and **25** [83%; $\delta_{C(=O)H} = 9.31$ and 9.49 (2 s)], respectively. Subsequently, these dialdehydes were pinacol-coupled under our previously described condi-

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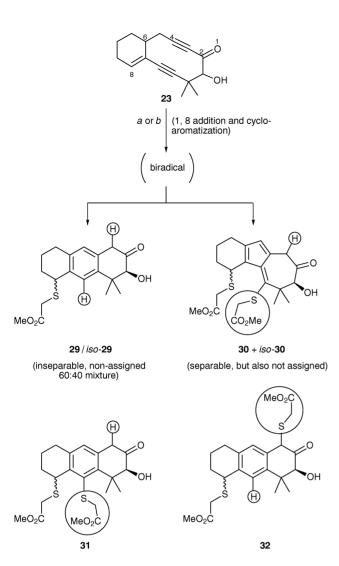
Scheme 3 (a) Ref. 15: Bu^tLi (1.1 equiv.), THF, -78 °C, 10 min; Tf₂O (1.1 equiv.), 78 °C, 15 min; 57%. (b) Ref. 15: LiHMDS (1.1 equiv.), THF, -78 °C, 1 h; Tf₂O (1.1 equiv.), -78 °C, 20 min; 74%. (c) 2,2-Dimethyl-3-butyn-1-ol (1.2 equiv.), Pd(PPh₃)₄ (5 mol %), CuI (10 mol %), piperidine (2.0 equiv.), THF, room temp., 3 h; 75%. (d) 2,2-Dimethyl-3-butyn-1-ol (1.1 equiv.), PdCl₂(PPh₃)₂ (5 mol %), CuI (10 mol %), Et₂O: Pr₂NH (3:1), 0°C, 3 h; addition of propargyl alcohol (1.0 equiv.), 0°C, overnight; room temp., 10 h; 80% (slightly contaminated). (e) Propargyl alcohol (1.1 equiv.), Pd(PPH₃)₄ (5 mol %), CuI (10 mol %), piperidine (2.0 equiv.), room temp., overnight; 83%. (f) Dess-Martin periodinane (2.2 equiv.), CH₂Cl₂, room temp., 30 min; 68% of a 85:15 mixture with the presumed E isomer. (g) 'Low-valent Ti' from TiCl₃ (18 equiv.) and Zn/Cu couple (55 equiv.), DME, -45°C,, 5.5 h; 56%. (h) ButOMgBr from EtMgBr solution in THF (3.0 equiv.) and ButOH (3.0 equiv.), azodicarbonyldipiperidide (1.2 equiv.), THF, 0 °C, 30 min; 55%

tions, $^{22d-f}$ *i.e.* with 'low-valent titanium'²⁵ generated from TiCl₃ and the Zn/Cu couple²⁶ in DME. Working at $-45\,^{\circ}$ C avoided the potentially competing^{22d-f} McMurry olefination.²⁷ The reduction products on which we concentrated, because they were readily isolable without contaminants were the bicyclic dienediynediols **21** [56%; $\delta_{\text{CH(OH)}} = 3.64$ (d, $J_{vic} = 8.6$ Hz) and 4.41 (d, $J_{vic} = 8.3$ Hz)] and **26** [80%; $\delta_{\text{CH(OH)}} = 3.64$ (d, $J_{vic} = 8.7$ Hz) and 4.40 (d, $J_{vic} = 8.6$ Hz)]. They were pure diastereomers according to the appearance of *single* δ_{H} and δ_{C} sets in the 300 MHz ¹H and 75 MHz ¹³C NMR spectra. Their configurations are suspected to be *cis* but are unestablished.

Scheme 4 (a) Propargyl alcohol (1.1 equiv.), $PdCl_2(PPh_3)_2$ (5 mol %), CuI (10 mol %), $Et_2O-Pr_2^iNH$ (3:1), room temp., 3 h; addition of 2,2-dimethyl-3-butyn-1-ol (1.0 equiv.), room temp., overnight; 39% pure 24 or 62% 24 containing typically 8 rel % of 'regioisomer' 19. (b) Same as (f) in Scheme 3; 83% of isomerically pure material. (c) Same as (g) in Scheme 3; 80%. (d) Same as (h) in Scheme 3; 76%

We had envisaged to finish the dienediyne ketone syntheses of Schemes 3 and 4 by an oxidation of the dienediynediols 21 and 26. However, CrvI reagents (PCC, PDC, Collins' reagent) or the Dess-Martin periodinane effected glycol cleavages in dienediynediols 21 or 26. Thus, their precursors, i.e. the dienediyne dialdehydes 20 and 25, respectively, were obtained again. Swern's oxidation conditions destroyed dienediynediols 21 and 26. Finally, we tried Narasaka et al.'s version²⁸ of the Oppenauer oxidation. It is basically a hydride abstraction by azodicarboxylic acid dipiperidide from the magnesium salt of the alcohol in question. Applied to our diols 21 and 26, this method brought about a chemoselective monooxidation in both instances. The propargylic OH group reacted while the electronically unactivated and sterically hindered OH group remained untouched. Thus, the ketones 23 (55%) and 28 (76%) were obtained.

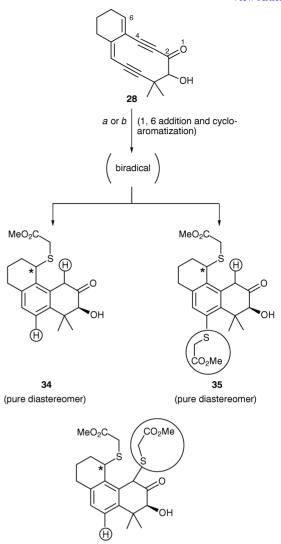
The dienediyne ketones 23 and 28 are acyloins. They can be distinguished from their isomers 22 and 27, respectively, through their 300 MHz 1 H NMR spectra. The shifts of the protons at the C=C bonds protruding from or lying within the 6-membered ring of acyloin 23 [$\delta_{C=CH_{exocyclic}} = 5.47$ (m_c); $\delta_{C=CH_{endocyclic}} = 6.45$ (td, $J_{vic} = 4.7$ Hz, $^5J = 1.5$ Hz)] resemble more those of the analogous protons in dialdehyde 20 [$\delta_{C=CH_{exocyclic}} = 5.56$ (br s); $\delta_{C=CH_{endocyclic}} = 6.51$ (td, $J_{vic} = 4.4$ Hz, $^5J = 1.2$ Hz)] than those of the diol 21 [$\delta_{C=CH_{exocyclic}} = 5.25$ (br s); $\delta_{C=CH_{endocyclic}} = 6.20$ (br t, $J_{vic} = 4.5$ Hz)]. This signifies that in acyloin 23 both C=CH groups are remotely conjugated with the C=O bond, like they are in 20. This supports the assignment of structure 23 at the expense of the alternative 22.



Scheme 5 (a) Methyl thioglycolate (2.1 equiv.), NEt₃ (1.0 equiv.), C_6H_6 , room temp., 10 h; 60: 40 **29**/iso-**29** (27%), **30** (27%) and iso-**30** (7%). (b) Same as (a) but without 1,4-cyclohexadiene; 60: 40 **29**/iso-**29** (16%), **30** (23%) and iso-**30** (16%). Entities taken up by the biradical intermediates to give the observed products or the indicated unobserved alternative structures are encircled

To the acyloin of Scheme 4 we assign structure 28. This seems justified even if the proton at the C=C bond protruding from the 6-membered ring $[\delta_{C=CH_{exocyclic}} = 5.35$ (td, ${}^4J_{9,11} \approx$ $^{5}J_{9,14} \approx 1.2 \text{ Hz}$] is characterized by the shift average of the analogous protons in dialdehyde **25** [$\delta_{C=CH_{exocyclic}} = 5.52$ (d, ${}^5J = 0.7$ Hz)] and in diol **26** [$\delta_{C=CH_{exocyclic}} = 5.20$ (br s)] and thus leaves open the question whether this C=C bond is conjugated with or isolated from the C=O group. Clarity comes from the proton at the C=C bond embedded in the 6membered ring of acyloin 28 [$\delta_{C=CH_{endocyclic}} = 6.59$ (td, $J_{vic} =$ 4.5 Hz, ${}^{5}J = 1.5$ Hz)]. The chemical shift resembles that of the corresponding proton in dialdehyde 25 $[\delta_{C=CH_{endocyclic}} = 6.70]$ (td, $J_{\text{vic}} = 4.5$ Hz, ${}^5J = 1.4$ Hz)] and not in diol 26 $[\delta_{C=CH_{endocyclic}}=6.22 \text{ (t, } J_{vic}=4.4 \text{ Hz)}].$ This resemblance and discrepancy mean that the acyloin of Scheme 4 contains the same substructure $HC=C-C\equiv C-C(=O)$ as the dialdehyde 25 and is devoid of the substructure HC=C-C=C-CH(OH) of the diol 26. Since formula 28 contains this substructure and formula 27 does not, the former is correct.

The newly prepared dienediyne ketones 23 and 28 cyclized/cyloaromatized when we treated them at room temperature with methyl thioglycolate and triethylamine for 2–10 h (Schemes 5, 6). Whether 1,4-cyclohexadiene was present or not at the same time affected the yields and product compositions only marginally. Each of the mentioned dienediyne ketones



Scheme 6 (a) Methyl thioglycolate (2.1 equiv.), NEt₃(1.0 equiv.), 1,4-cyclohexadiene (23 equiv.), CH₂Cl₂, room temp., 2 h; 34 (38%) and 35 (16%). (b) Methyl thioglycate (20 equiv.), NEt₃ (1.0 equiv.), CH₂Cl₂, room temp., 10 h; 34 (18%) and 35 (30%). Entities taken up by the biradical intermediate to give the observed products or the indicated unobserved alternative structure are encircled

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afforded two types of products. Their combined yields reached 61% starting from 23 and 54% starting from 28; these numbers match the highest values reported in the literature systems^{6,8–15} (64% for **10d**,¹¹ 56% for **10b**¹⁰). One product type corresponds to the uptake of one equivalent of methyl thioglycolate and one equivalent of hydrogen; the other is characterized by the uptake of two equivalents of methyl thioglycolate. Specifically, the dienediyne ketone 23 and methyl thioglycolate provided up to 27% type-1 product (29/iso-29) as an unseparable 60:40 mixture of unassigned diastereomers or up to 39% type-2 products (30/iso-30) as separable major and minor diastereomers, respectively. Similarly, the dienediyne ketone 28 and methyl thioglycolate gave to up to 38% type-1 product (34) or up to 30% type-2 product (35), both of which were single diastereomers. Product 35 is distinguishable from the isomeric structure 36 by the presence of one instead of two sp²-bound protons.

The structures of the cyclized/cycloaromatized products 29, iso-29, 30, iso-30, 34 and 35 were deduced from their NMR spectra unequivocally (29, iso-29, 34, 35; Tables 1 and 2) or represent, in the absence of crystal structure analyses, tentative assignments (30, iso-30; Table 1). The formation of pairs of diastereomers was inferred from the occurrence of two very

Table 1 Selected 300 MHz ¹H and 75.5 MHz ¹³C NMR data (CDCl₃) of the cycloaromatized/cyclized products obtained from ketone **23** (500 MHz ¹H- and 125.7 MHz ¹³C NMR for compounds **30** and *iso-30*) and, for comparison, of the known compound **33**. ^{9b} Coupling constants in Hz

Nucleus	29 / <i>iso</i> - 29	33	30	iso- 30
1-H	$4.30^a (t)/4.31^a (t)$	4.84 (br d)	4.41 (br s)	4.20 (m _c)
5-H ₂	$3.74 (J_{qem} \text{ unknown})$	3.63 (br d)	2.89 and 3.22 ($J_{gem} = 17.8$)	2.69 and 3.64 ($J_{gem} = 16.8$)
$7-H_2$	$4.20 \ (J = 3.7)/4.17 \ (J = 3.4)$	2.48 and 2.51 ($J_{qem} = 15.0$)	4.18 (J = 2.5)	$4.13 (J = 2.5)^b$
9-H (s)	$7.43^{c}/7.42^{c}$	7.27	_	_
10-H	6.86^{c}	6.92	6.99	6.98
$\mathrm{S\text{-}C}H_{2}\;(J_{\mathit{gem}})$	3.23 and 3.36 ($J = 15.1$)/3.21 and 3.35 ($J = 14.9$)	2.75 and 3.09 ($J = 16.3$)	3.19 and 3.36 ($J = 14.5$)/3.66 and 3.70 ($J = 15.2$)	3.19 and 3.36 ($J = 14.7$)/3.65 and 3.71 ($J = 15.2$)
C-1	52.37/52.39	d	41.31	41.08
C-6	209.74	d	198.98	199.06
C-7	80.22/80.32	d	80.21	80.66
CO_2CH_3	171.16/171.11	d	169.96 and 170.72	169.94 and 170.64

^a Assignment exchangeable. ^b Assignment exchangeable. ^c Assignment exchangeable. ^d No ¹³C NMR spectrum of this compound has been published.

Table 2 Selected ¹H and ¹³C NMR data (CDCl₃) of the cycloaromatized products (500 MHz/125.7 MHz) obtained from ketone **28** and, for comparison, of the known compounds **37** (300 MHz/75.5 MHz), ¹⁵ **38** (500 MHz/125.7 MHz; C₆D₆), ²⁹ **39** (400 MHz) ¹⁴ and **40** (400 MHz), ¹⁴ Coupling constants in Hz

Nucleus	34	37	38	39	35	40
4-H	4.12 (br s)	4.18 (br s)	4.02	4.40 (br d)	4.12 (br s)	4.41 (br d)
5-H ₂	3.67 and 4.29	3.50 and 4.00	3.44 and 4.43	3.52 and 4.07	3.65 and 4.28	3.48 and 4.04
	$(J_{gem} = 22.1)$ 4.23	$(J_{gem} = 20.4)$	$(J_{gem} = 21.7)$	$(J_{gem} = 22.0)$	$(J_{gem} = 22.1)$	$(J_{gem} = 22.0)$
$7-H_{(2)}$	4.23	2.56 or 3.03	2.22	2.50 and 2.58	4.21	2.49 and 2.57
				$(J_{gem} = 14.5)$ 7.31 ^d		$(J_{gem} = 14.8)$
9-H	7.30^{a}	7.05^{b}	7.03^{c}		_	_
	$(d, J_{9,10} = 8.1)$	$(d, J_{9,10} = 7.9)$	$(d, J_{9,10} = 8.0)$	$(d, J_{9,10} = 8.0)$		
10-H	7.07^a	6.95^{b}	6.81^{c}	7.15^{d}	7.36 (s)	7.36 (s)
	$(d, J_{10,9} = 8.1)$	$(d, J_{10,9} = 7.9)$	$(d, J_{10,9} = 8.3)$	$(d, J_{10,9} = 7.9)$		
$S-CH_2$	3.20 and 3.35	3.19 and 3.36	2.73 and 2.93	3.25 and 3.32	3.19 and 3.34	3.24, 3.32, 3.63 and 3.66
$^{(J_{\it gem})}_{ ext{C-4}}$	(J = 14.4)	(J = 14.4)	(J = 14.1)	(J = 14.8)	$(J = 14.4), 3.67 (2 \times)$	(J = 14.6 and 14.8)
	42.39	41.80	42.34	e	42.51	J
C-6	209.65	210.75 or 210.76	207.60	e	209.33	J
C-7	79.73	38.41 or 40.54	53.61	e	79.49	f
CO_2CH_3	170.73	170.80	170.14	e	170.03 and 170.67	J

^a Assignment interchangeable. ^b Assignment interchangeable. ^c Assignment interchangeable. ^d Assignment interchangeable. ^e No ¹³C NMR spectrum of this compound has been published. ^f No ¹³C NMR spectrum of this compound has been published.

similar ¹H and ¹³C NMR data sets for the compound pairs 29/iso-29 and 30/iso-30. Conversely, the most likely interpretation for observing single ¹H and ¹³C NMR data sets in the compounds 34 and 35 is that each of them is diastereomerically pure.

Let us first consider pertinent NMR data of the safely assignable cyclization/cycloaromatization products. Compounds 29 and iso-29 are octahydrophenanthrenones (Table 1). They exhibit singlets for their para-positioned aromatic

protons. The resonances of 5-H_2 in 29 and iso-29 resemble the 5-H_2 shifts in the related compound 33.9^{9} Moreover, the aliphatic proton signals of 29 and iso-29 in general resemble those of the newly prepared octahydroanthracenones 34 and 35 (Table 2). The latter two compounds, in turn, resemble the aliphatic ^{1}H NMR resonances of their deoxygenated analogues 37^{15} and 38^{29} or their deoxygenated/demethylated analogues 39^{14} and 40^{14} (Table 2). These analogies in the ^{1}H NMR data are paralleled by a reliable similarity of the ^{13}C

NMR shifts of all carbon centers that are analogously substituted. Naturally, this statement can be probed only as far as these ¹³C NMR shifts are available; they are unknown for the reference compounds 33, ^{9b} 39, ¹⁴ and 40. ¹⁴

Let us now turn to the structures of compounds 30 and iso-30 obtained from the dienediyne ketone 23 through the net

uptake of two equivalents of methyl thioglycolate. Contemplating the ¹H and ¹³C NMR spectra of each of these species (Table 1) separately one can take them for the diastereomeric octahydroanthracenones 31 depicted in Scheme 5. Clearly, these compounds were not the isomers 32 because they contain one sp²-bound proton and not two. However, upon

closer scrutiny one doubts that the structures 31 are correct. There are two singularities in juxtaposing the NMR data in question and the NMR data of the reference compounds 29, iso-29, 33-35 or 37-40 (Tables 1, 2). (a) The geminal coupling constant between the diastereotopic protons 5-H₂ of 30 and iso-30 measures 17.8 and 16.8 Hz, respectively; this is different from the reference values of $20.4 \leqslant J_{gem} \leqslant 22.1$ Hz for benzylic 5-H₂ groups. (b) The keto group of 30 and iso-30 resonates 10 ppm upfield from the standard value of 210 ± 1 ppm encountered elsewhere. As a consequence we tend to discard the octahydrophenanthrenone structure 31 and tentatively suggest in place of them the octahydrobenzoazulenone structures shown. These species would arise from a Schmittel cyclization of the enyneallenyl ketone 44 (formula: Scheme 7) followed by a regioselective addition of methyl thioglycolate to the resulting fulvene biradical. We are unaware of related octahydrobenzoazulenones and the chemical shift of their C=O groups. Therefore, we are not able to substantiate the validity of the proposed structural assignments.

Discussion

The structurally unambiguous cycloaromatization products 29 and iso-29 are proposed to arise from the Saito-Myers biradical 46 (Scheme 7). They are formed through a two-step 1,6-addition of methyl thioglycolate to the C=C−C=C−C=C motif of the dienediyne ketone 23, the addition product being the enyneallenyl ketone 44. Biradical 46 is a toluene biradical, i.e. a species that contains, at a qualitative level of consideration, one localized and one delocalized radical center. It gives the valence-saturated species 29 and its diastereomer iso-29 through the uptake of two hydrogen atoms. They stem from methyl thioglycolate and/or from 1,4-cyclohexadiene.

The cycloaromatizations starting from the dienediyne ketones 7 and 9-14 of Scheme 2 could a priori also proceed by a neocarzinostatin-type cycloaromatization mechanism, i.e. by an enyne[3]cumulene → styrene biradical conversion.³a If this alternative mechanism applied, a two-step 1,8-addition of methyl thioglycolate to the C=C-C=C-C=C-C=O moiety of ketone 23 would be the starting point of our reaction and lead to the enyne[3]cumulenol 45 (Scheme 7). Its neocarzinostatin-type cycloaromatization would give the styrene biradical 47. The latter could either isomerize to the Saito-Myers biradical 46 and continue to react as already described. Or, it could itself pick-up two hydrogen atoms from methyl thioglycolate and/or from 1,4-cyclohexadiene. Diastereomeric enols would form whose ketone tautomers are identical with the observed cycloaromatization products 29 and iso-29.

The styrene biradical 47 is a species with two nonconjugated radical centers. Therefore, a styrene biradical like 47 should be less stable and probably also less rapidly formed than an isomeric toluene biradical like 46. One would expect an exception to this generalization only if a ring-size effect destabilizes the styrene biradical less than the toluene biradical. However, such an exception can be excluded for the case at hand by density functional theory, as described in the subsequent part of this paper. These calculations do not only suggest that compounds 29 and iso-29 form only via the Saito-Myers pathway but also that the Saito-Myers mechanism prevails for the cycloaromatization of dienediyne ketones 10-13 in which the C=O bond lies in a 10-membered ring.

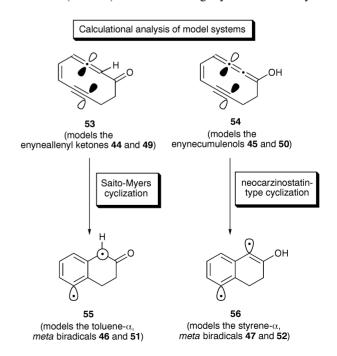
Having thus established that the biradical precursor of the cycloaromatization of Scheme 7 is 46, one can now interpret the chemoselectivity of its quenching reaction. As Scheme 5 shows, we obtained the hydrogen addition products 29 and iso-29 and none of the thiol addition products 31 or 32. This selectivity can be rationalized as follows. (a) The phenyl radical moiety of biradical 46 is sterically hindered through two ortho substituents, one of which is additionally once and

the other twice branched. Accordingly, this radical center cannot combine as fast with a MeO₂C-CH₂-S· radical, even if the latter is formed close by, as it abstracts an H atom from methyl thioglycolate or 1,4-cyclohexadiene. (b) The benzyl radical moiety of biradical 46 is considerably more stable than the phenyl radical moiety. Hence, it, too, scavenges MeO₂C-CH₂-S· radicals only slowly. It is long-lived enough to be saturated by an H-atom transfer from methyl thioglycolate or from 1,4-cyclohexadiene (to give 29/iso-29, q. e. d.).

The other genuine cycloaromatization products 34 and 35 of the present study arose from the dienediyne ketone 28. For compounds 34 and 35, too, there is the dichotomy of a Saito-Myers or a neocarzinostatin-type formation mechanism (Scheme 8). For the reasons given in the discussion of Scheme 7, in Scheme 8 the Saito-Myers mechanism (two-step 1,6addition of methyl thioglycolate → enyneallenyl ketone $49 \rightarrow \text{toluene biradical } 51 \rightarrow 34/35)$ should again be preferred over the neocarzinostatin pathway (two-step 1,8-addition of methyl thioglycolate \rightarrow enynecumulenol 50 \rightarrow styrene biradical $52 \rightarrow 34/35$ or enol form of 34/35). In light of this insight, comparing the chemoselectivities of the quenching reactions of the most likely biradical intermediates 51 (Scheme 8) and 46 (Scheme 7) is instructive. In both instances the benzyl radical moiety reacts only with methyl thioglycolate or 1,4-cyclohexadiene and not with MeO₂C-CH₂-S· (which would have furnished the non-observed quenching product 36); a plausible reason was given in the context of Scheme 7. The phenyl radical moiety is less sterically hindered in biradical 51 than in biradical 46; consequently, other than 46, 51 has a choice of combining with a MeO₂C-CH₂-S· radical (to give 35) or abstracting an H atom from methyl thioglycolate or 1,4-cyclohexadiene (to give 34).

Calculations

In the preceding discussion it was pointed out that there are two conceivable reaction paths, namely the Saito-Myers and the neocarzinostatin-type cyclizations, by which methyl thioglycolate addition products of the dienediyne ketone 23 (i.e., the enyneallenyl ketone 44 or the tautomeric enynecumulenol 45; Scheme 7) or of the dienediyne ketone 28 (i.e., the enyneallenyl ketone 49 or the tautomeric enynecumulenol 50; Scheme 8) could lead to the aromatics 29/iso-29 (from 23) or 34 and 35 (from 28). The same ambiguity concerns the cyclo-



Scheme 9

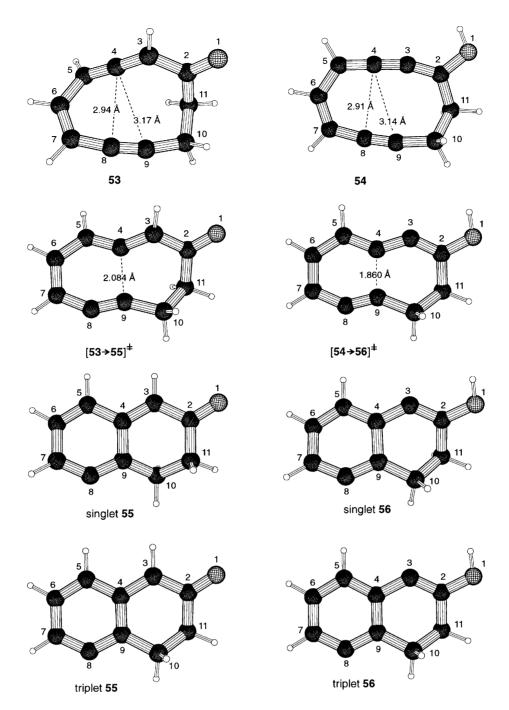


Fig. 1 Geometries of reactants 53, 54 and transition states $[53 \rightarrow 55]^{\pm}$, $[54 \rightarrow 56]^{\pm}$ calculated at the B3LYP/6-31G* level. Geometries of the products: singlet 55 and singlet 56 [CAS(2,2)/6-31G] and triplet 55 and triplet 56 (UB3LYP/6-31G*)

aromatization mechanism of all previously cycloaromatized dienediyne ketones (7, 9, 10a-e, 11-14). A mechanistic clarification seems therefore important. We provide it here by a density functional theory treatment of the most abundant subclass of these reactions where a ten-membered ring core structure participates. Specifically, we investigated the energy hypersurfaces of the Saito-Myers reaction $53 \rightarrow 55$ and the neocarzinostatin-type cycloaromatization $54 \rightarrow 56$ (Scheme 9). Both reactions describe a transition from a closed-shell to an open-shell hypersurface. Therefore, different methods had to be employed for computationally treating the starting materials, transition structures and products (methodological and computational details: captions of Fig. 1 and 2). Selected bond lengths and bond angles of reactants, transitions states and products are given in Table 3 (geometries: Fig. 1). Fig. 2 depicts the energies of the stationary points on the hypersurface of both reactions.

According to our density functional theory calculations

 $(B3LYP/6-31G*^{30,31})$ the allenyl ketone 53 is 21.1 kcal mol⁻¹ more stable than the tautomeric cumulenol 54. The cycloaromatization product, toluene singlet biradical 55, is thermodynamically favored over the tautomeric styrene singlet biradical 56 by as much as 35.3 kcal mol⁻¹, according to ab initio calculations [CASMP2(2,2)/6-31G//CAS(2,2)/6.31-G].32

Biradicals 55 and 56 differ fundamentally in their electronic structure. In biradical 56 the two 'radical centers' interact strongly with one another mainly by through-bond conjugation. The orbital degeneracy that would be expected for a 'pure' biradical is thereby cancelled in the following way. One of the two in-plane p-('radical') orbital combinations—which would be weakly (!) bonding through space—is raised in energy, the other-which would be weakly (!) antibonding through space—is somewhat lowered. The biradical character of a species can be expressed by the occupancy of both active MOs in a two-orbital two-electron multireference description. According to our CAS(2,2) calculations there are 0.77 and 1.23

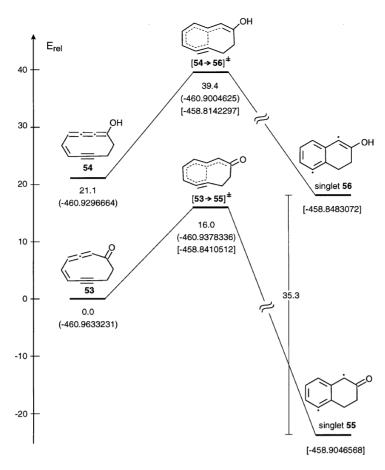


Fig. 2 Calculated relative energies of the stationary points of the Saito-Myers reaction $53 \rightarrow 55$ and the neocarzinostatin-type cyclization $54 \rightarrow 56$. Absolute energies (a.u.) by B3LYP/6-31G*/B3LYP/6-31G* in parentheses (), by CASMP2(2,2)/6-31G/CAS(2,2)/6-31G in brackets []. Relative energies in kcal mol⁻¹ without parentheses; energies of cumulenol 54 and of transition states are relative to allenyl ketone 53 at the B3LYP level; energy differences between biradical products 55 and 56 calculated at the CAS level

electrons in the through-bond bonding and antibonding inplane p-orbital combination, respectively (both of them are mainly located at the 'radical centers'). This is about the same degree of biradical character as o-didehydrobenzene, in which the occupancies of the respective MOs are 0.80 and 1.20.

In biradical 55 the unpaired electrons do not interact with one another on (approximate) symmetry grounds. They occupy orbitals that are almost orthogonal. One is mainly located at the dehydro position of the benzene ring, *i.e.* inplane. The second is delocalized over the π system and

exhibits the largest coefficient in the α position with respect to the carbonyl group. The orbital occupancy of both active orbitals is 1.06 and 0.94, which indicates an almost pure biradical state.

The activation barrier of the Saito–Myers cyclization $53 \rightarrow 55$ (16.0 kcal mol⁻¹) is slightly lower than that of the neocarzinostatin-type cyclization $54 \rightarrow 56$ (18.3 kcal mol⁻¹) (B3LYP/6-31G*). More importantly, the transition state $[53 \rightarrow 55]^{\neq}$ of the Saito–Myers cyclization lies 23.4 kcal mol⁻¹ below the transition state $[54 \rightarrow 56]^{\neq}$ of the

Table 3 Selected bond distances $[\mathring{A}]$ and bond angles $[\mathring{\circ}]$ of 53, 54, singlet 55, triplet 55, singlet 56, triplet 56 and the transition states $[53 \rightarrow 55]^{\neq}$ and $[54 \rightarrow 56]^{\neq}$. The numbering of atoms is shown in Fig. 1 and does not follow IUPAC nomenclature

	53 ^a	54 ^a	singl. 55^b	tripl. 55 ^a	singl. 56 ^b	tripl. 56 ^a	$[53 \rightarrow 55]^{\neq a}$	$[54 \rightarrow 56]^{\neq a}$
1-2	1.222	1.368	1.227	1.234	1.371	1.363	1.225	1.364
2-3	1.493	1.333	1.453	1.450	1.381	1.332	1.469	1.333
3-4	1.315	1.263	1.449	1.422	1.449	1.441	1.346	1.338
4-5	1.317	1.338	1.399	1.420	1.390	1.404	1.362	1.375
5-6	1.469	1.452	1.386	1.387	1.391	1.396	1.424	1.418
6-7	1.356	1.363	1.393	1.410	1.388	1.404	1.379	1.378
7-8	1.417	1.415	1.375	1.381	1.379	1.381	1.394	1.397
8-9	1.214	1.215	1.374	1.370	1.367	1.374	1.252	1.272
9-10	1.459	1.464	1.511	1.513	1.516	1.519	1.467	1.497
4-9	3.173	3.139	1.409	1.438	1.417	1.428	2.084	1.860
4-8	2.936	2.905	_	_	_	_	_	_
2-3-4	127.7	163.6	121.6	122.9	126.4	126.9	125.9	135.1
3-4-5	174.8	178.0	121.3	121.5	124.1	124.4	142.4	139.6
7-8-9	169.9	171.8	125.7	126.9	126.2	126.6	145.2	141.0
8-9-10	166.6	169.5	123.4	124.6	124.5	125.0	153.3	143.4
^a B3LYP/6-31G*. ^b CAS(2,2)/6-31G.								

neocarzinostatin-type cyclization (B3LYP/6-31G*). Since under the reaction conditions, *i.e.* in the presence of NEt₃, the tautomerism $54 \rightarrow 53$ should be faster than the two cycloaromatizations in question, the considerably greater stability of the Saito–Myers vs. the neocarzinostatin-type transition state means that the far preferred cycloaromatization in this system is the Saito–Myers reaction.

Scrutinizing the structures (Table 3, Fig. 1) of the reactants, transition states and products of these cycloaromatizations one notes large deviations from linearity at the sp-hybridized carbon atoms of the reactants 53 and 54. They indicate high ring strain. The transannular distance of the carbon atoms between which the C-C single bond forms in the course of the cycloaromatization is similar in 53 (3.173 Å) and 54 (3.139 Å). It is, of course, shortened considerably in the corresponding transition states $[53 \rightarrow 55]^{\neq}$ and $[54 \rightarrow 56]^{\neq}$, respectively, but interestingly not to the same extent. In $[53 \rightarrow 55]^{\neq}$ it measures 2.084 Å as opposed to 1.860 Å in $[54 \rightarrow 56]^{\neq}$. This bond length difference is in agreement with the Hammond postulate. The exothermic Saito-Myers cyclization occurs over a more reactant-like transition state with a still long $C \cdots C$ distance and the almost thermoneutral neocarzinostatin-type cyclization occurs over a less reactant-like transition state with an already shorter $C \cdot \cdot \cdot C$ distance.

Experimental

Methodological and computational details

All calculations were performed using the implementation of the B3LYP functional, 30,31 the CASSCF and CASMP2 procedure and the standard 6-31G and 6-31G* basis set available in the Gaussian94 package.³² The CASSCF active space for the treatment of the singlet diradicals 55 and 56 comprised two electrons distributed in two orbitals. Because both species are not perfectly planar, only approximate symmetry considerations could be used for the selection of the active orbitals. For 55 a CAS(UNO)33 calculation was performed and the highest occupied σ -type MO with a very large coefficient at C-8 (for numbering see Fig. 1) and the singly occupied π -type orbital were selected for the CASSCF treatment. For 56 the two σ-type orbitals with coefficients mainly located at the 'diradical centers' C-3 and C-8 were included in the twoelectron two-orbital treatment. During optimization of 55 the orbital ordering had to be readjusted several times. The multiconfiguration approach was used in order to take into account the large nondynamical correlation effects, especially in 55. However, it is known that dynamical correlation has to be included in multiconfigurational treatments to obtain results that are quantitatively or even qualitatively reliable.³⁴ We performed CASMP235,36 single-point calculations at CAS(2,2)/6-31G geometry, including second-order Møller-Plesset perturbation theory,³⁷ to provide dynamic correlation (between electrons considered in the active space and the remaining electrons) for the CAS wavefunction. Both transition states $[53 \rightarrow 55]^{\neq}$ and $[54 \rightarrow 56]^{\neq}$ were also calculated at the above level [CASMP2(2,2)/6-31G//CAS(2,2)/6-31G]. The low occupancy of the higher orbital (diagonal entries in the symbolic CAS density matrix: 1.91, 0.09 for $[53 \rightarrow 55]^{\neq}$ and 1.92, 0.08 for $[54 \rightarrow 56]^{\neq}$) accounts for the low diradical character of the transition states. The density functional theory calculations at the B3LYP/6-31G* level, therefore, should provide an estimated value for the activation enthalpies for ring closure. A large diradical character is developed only at a later stage of the reaction coordinate. The occupancies of the two active orbitals are 1.06, 0.94 for 55, which corresponds to an almost pure diradical, and 1.23, 0.77 for 56. Benchmark calculations using the above levels on the parent system of the Bergman cyclization are in very good agreement with experimental data: 38 ΔH^{\ddagger} (cyclization) = 31.0 kcal mol $^{-1}$ (B3LYP/6-31G*), 28.2 kcal mol $^{-1}$ (expt.), ΔH^{\ddagger} (ring opening) = 19.7 kcal mol $^{-1}$ [CASMP2(6-31G)/CAS(2,2)/6-31G], 21.6 kcal mol $^{-1}$ (expt.). For optimizing the structures of 53, 54, singlet 55, triplet 55, singlet 56, triplet 56, and the transition states $[53 \rightarrow 55]^{\ddagger}$ and $[54 \rightarrow 56]^{\ddagger}$ we used the B3LYP hybrid functional 30,31 and the 6-31G* basis set. All stationary points were characterized by harmonic frequency analysis.

Synthetic details

All reactions were performed in oven-dried (100 °C) glassware under N2. THF was freshly distilled from K and CH2Cl2 from CaH2. Products were purified by flash chromatography²³ on Merck silica gel 60 (eluents given in brackets). Yields refer to analytically pure samples. Isomer ratios were derived from suitable ¹H NMR integrals. ¹H [CHCl₃ (7.26 ppm) as internal standard in CDCl₃ or C₆HD₅ (7.16 ppm) as internal standard in C_6D_6] and ^{13}C [CDCl₃ (77.00 ppm) as internal standard in CDCl₃ or C₆D₆ (128.00 ppm) as internal standard in C₆D₆] NMR spectra were acquired on Varian VXR 200, Bruker AMX 300, and Varian VXR 500S spectrometers; integrals are in accord with assignments; coupling constants in Hz; APT ¹³C NMR spectra: "+" for CH or CH₃, "-" for CH₂ or C_{quat}. The assignments of ¹H and ¹³C NMR resonances refer to the IUPAC nomenclature with primed numbers belonging to the side-chain(s) in the order of their IUPAC appearance in the name. Combustion analyses: M. Beller, Institute of Organic Chemistry, University of Göttingen. MS: Dr. G. Remberg, Institute of Organic Chemistry, University of Göttingen. IR spectra: Perkin-Elmer 1600 Series FTIR as CDCl₃ solution in a NaCl cuvette.

Z-[2-(3,3-Dimethyl-4-hydroxy-1-butynyl)-2-cyclohexenylidene | methyl trifluoromethanesulfonate (18). At room temperature, we first added a solution of 2,2-dimethyl-3-butyn-1ol (634 mg, 6.44 mmol, 1.2 equiv.) in THF (6 mL) and then CuI (102.0 mg, 0.536 mmol, 10 mol %) to the bistriflate 17 (2.094 g, 5.36 mmol), Pd(PPh₃)₄ (309.6 mg, 0.268 mmol, 5 mol %) and piperidine (1.10 μ L, 914 mg, 10.7 mmol, 2.0 equiv.) in THF (34 mL). The mixture was stirred for 3 h and then hydrolyzed with brine (20 mL). After extraction with ether, the organic layer was dried over MgSO4. The solvent was removed in vacuo at 0 °C. Flash chromatography of the residue (pentane-ether, 2:1) afforded 18 (1.295 g, 75%). IR $(CDCl_3)$: v = 3585, 3435, 3100, 2975, 2870, 2250, 1730, 1655, 1415, 1210, 1135, 1115, 1055, 1015, 915 cm⁻¹. ¹H NMR (200 MHz, CDCl₃): $\delta = 1.22$ [s, 3"-(CH₃)₂], 1.68–1.78 (m, 5'-H₂), 2.10–2.23 (m, 6'-H₂), 2.27 (dt, $J_{4',3'} = J_{4',5'} = 4'$ -H₂), 3.43 (s, 4"-H₂), 3.70 (br s, OH), 6.34 (td with strong roof effect towards lower field, $J_{3',4'} = 4.4$, ${}^5J_{3',1} = 1.6$, 3'-H), 6.38 (s, 1-H). 13 C NMR (75.5 MHz, CDCl₃): $\delta = "-"21.82$ (C-5'), "+"25.14 [3-(CH₃)₂], "-"26.50 and "-"27.54 (C-4', C-6'), "-"34.76 (C-3"), "-"71.46 (C-4"), "-"79.70 and "-"94.05 (C-1", C-2"), "-"115.87 and "-"124.20 (C-1', C-2'), "-"118.64 (q, ${}^{1}J_{C,F} = 321.4$, CF₃), "+"129.10 (C-1), "+"143.22 (C-3'). $C_{14}H_{17}O_4F_3S$ [M⁺]: calcd. 338.0799; the exact molecular mass (± 2 ppm; R = 10000) was confirmed by EI HRMS (70

Z-4-[2-(3,3-Dimethyl-4-hydroxy-1-butynyl)-2-cyclohexenylidene]-2-butyn-1-ol (19). Procedure A: To a solution of the monocoupling product 18 (1.295 g, 4.02 mmol), Pd(PPh₃)₄ (232.2 mg, 0.201 mmol, 5 mol %) and CuI (76.6 mg, 0.402 mmol, 10 mol %) in THF (15 mL) was first added a solution of propargyl alcohol (451.1 mg, 8.04 mmol, 1.1 equiv.) in ether (5 mL) and then piperidine (0.80 mL, 69 mg, 8.0 mmol, 2.0 equiv.). The mixture was stirred overnight at room temperature and then hydrolyzed with brine (30 mL). After

extraction with ether, the organic layer was dried over MgSO₄. The solvent was removed in vacuo at 0°C. Flash chromatography of the residue (pentane-ether, 1:1) afforded 19 (834.4 mg, 83%). Procedure B: A solution of the bistriflate 17 (1.179 g, 3.02 mmol), PdCl₂(PPh₃)₂ (106.0 mg, 0.151 mmol., 5 mol %) and CuI (57.5 mg, 0.302 mmol, 10 mol %) in ether (15 mL) was cooled to 0 °C. A solution of 2,2-dimethyl-3butyn-1-ol (327.1 mg, 3.32 mmol, 1.1 equiv.) in ether (15 mL) and Pr₂NH (10 mL) were added in this order. The mixture was stirred for 3 h at this temperature. Propargyl alcohol (169.4 mg, 3.02 mmol, 1.0 equiv.) dissolved in ether-Pr₂NH (3:1; 8 mL) was added. The resulting mixture was stirred overnight at 0 °C and for 10 h at room temperature and then hydrolyzed with brine (30 mL). After extraction with ether, the organic layer was dried over MgSO₄. The solvent was removed in vacuo at 0°C. Flash chromatography of the residue (pentane-ether, 1:1) afforded 19 (590.0 mg, 80%) admixed with some of the 'regioisomer' 24. 19 could be liberated from 24 by keeping the early fractions and rechromatographying several times the late fractions of the preceding separation. IR (CDCl₃): v = 3505, 3155, 2970, 2935, 2870, 2255, 1795, 1650, 1470, 1380, 1165, 1095, 1050, 1015, 900 cm $^{-1}$. ^{1}H NMR (200 MHz, CDCl $_{3}$): $\delta = 1.25$ [s, 3"-(CH $_{3}$) $_{2}$], 1.68–1.77 (m, 5'-H₂), 2.22–2.32 (m, 4'-H₂, 6'-H₂), ca. 2.70 (br s which is only detectable in the integral, OH), 3.43 (s, 4"-H₂), 4.40 (d, ${}^{5}J_{1,4} = 2.7$, 1-H₂), 5.45 (almost not resolved d, ${}^{5}J_{4,1} = 1.1, 4\text{-H}$), 6.32 (td, $J_{3',4'} = 4.4, {}^{5}J_{3',4} = 1.4, 3'\text{-H}$). ${}^{13}\text{C}$ NMR [75.5 MHz, CDCl₃]: $\delta = \text{``-''} 22.42$ (C-5'), "+" 25.51 [3"-(CH₃)₂], "-"26.87 and "-"33.43 (C-4', C-6'), "-"35.23 (because of relatively low intensity: C-3"), "-"51.70 (C-1), "-"71.87 (C-4"), "-"81.30, "-"82.90, "-"94.27 and "-"94.91 (C-2, C-3, C-1", C-2"), "+"104.75 (C-4), "-"120.17 (C-2'), "+" 143.00 (C-3'), "-" 143.78 (C-1'). $C_{16}H_{20}O_2$ [M⁺]: calcd. 244.1463; the exact molecular mass (± 2 ppm; R = 10000) was confirmed by EI HRMS (70 eV).

Z-4-[2-(4-Oxo-3,3-dimethyl-1-butynyl)-2-cyclohexenylidene | -2-butyn-1-al (20): (contaminated with varying amounts of the presumed E isomer after chromatography). To a stirred solution of the Dess-Martin periodinane (1.593 g, 3.75 mmol, 2.2 equiv.) in CH₂Cl₂ (15 mL) a solution of diol **19** (417.2 mg, 1.71 mmol) in CH₂Cl₂ (10 mL) was added at room temperature. After 30 min the reaction mixture was diluted with ether (30 mL) and poured into a saturated aqueous solution of NaHCO₃ containing a sevenfold excess of Na₂S₂O₃. The mixture was stirred for ca. 10 min until the solid was dissolved. Then the layers were separated. The organic layer was washed with water and dried over MgSO₄. The solvent was removed in vacuo at 0 °C. Isolation by flash chromatography (pentane-ether) gave an unseparable mixture of compounds presumed to constitute a 85:15 mixture of the title compound and its E isomer (279.1 mg, 68%). The NMR spectra of the described and other mixtures obtained similarly allowed us to identify the resonances of 20. IR (CDCl₃): v = 2985, 2935, 2865, 2255, 2165, 1730, 1650, 1600, 1460, 1390, 1270, 1180, 1075 cm⁻¹. ¹H NMR (200 MHz, CDCl₃): $\delta = 1.36$ [s, 3"-(CH₃)₂], 1.73–1.82 (m, 5'-H₂), 2.32 (tdm_e, $J_{4',3'} \approx J_{4',5'} \approx 5.5$, 4'-H₂), 2.36–2.42 (m, 6'-H₂), 5.56 (br s, 4-H), 6.51 (td, $J_{3',4'}$ = 4.4, ${}^5J_{3',4}$ = 1.2), 9.28 (d, ${}^5J_{1,4}$ = 1.2, 1-H), 9.52 (s, 4"-H). 13 C NMR [75.5 MHz, CDCl₃; contains small peaks of contaminant(s)]: δ = "-"21.90 (C-5'), "+'22.41 (2"-CH₃), "-"26.79 and "-"33.41 (C-4', C-6'), "-"43.14 (C-2"), "-"82.33, "-"91.20, "-"93.83 and "-"95.02 (C-2, C-3, C-3", C-4"), "+" 102.41 (C-4), "-" 120.17 (C-2'), "+" 146.45 (C-3'), "-"151.21 (C-1'), "+"176.80 (C-1), "+"198.26 (C-1"). C₁₆H₁₆O₂ [M⁺]: calcd. 240.1150; the exact molecular mass $(\pm 2 \text{ ppm}; R = 10000)$ was confirmed by EI HRMS (70 eV).

4,4-Dimethylbicyclo [8.4.0] tetradeca-1(14),9-diene-2,7-diyne-5,6-diol (21): (single diastereomer of unknown configuration).

Cl₃Ti(DME)₂ was prepared by refluxing TiCl₃ (6.6 g, 42.8 mmol, 17.9 equiv.) in freshly distilled DME (200 mL) for 1.5 days. The Zn/Cu couple (8.5 g, 131.04 mmol, 54.6 equiv.) was added. The resulting mixture was refluxed for 5 h. A solution of the crude dialdehyde 20 (because of its presumed $Z \rightarrow E$ isomerization upon isolation, it was used crude in the following experiment, assuming that it was formed in 70% yield by the Dess-Martin oxidation of diol 19; 574 mg, 2.39 mmol) in DME (80 mL) was added at -45 °C by means of a syringe pump over a period of 5 h. After 30 min, the reaction mixture was diluted with ether and then hydrolyzed by adding a saturated aqueous solution of NaHCO₃ (50 mL). After filtration over celite, the aqueous layer was extracted with ether, the organic layer was washed with brine and dried over MgSO4. The solvent was removed in vacuo at 0°C. Flash chromatography (pentane-ether, 2:3) of the residue gave 21 (324.4 mg, 56%). IR (CDCl₃): v = 3560, 3415, 2975, 2935, 2870, 2250, 1730, 1465, 1385, 1260, 1180, 1060 cm⁻¹. ¹H NMR (200 MHz, CDCl₃; contains ether): $\delta = 1.24$ and 1.34 [2s, 4-(CH₃)₂], 1.64–1.74 (m, 12-H₂), 2.21 (td, $J_{13,12} = J_{13,14} = 5.5$, 13-H₂), $2.33 \text{ (tm}_{c}, J_{11,12} \approx 6.1, 11\text{-H}_{2}), 2.48 \text{ (br s, OH)}, 2.98 \text{ (br s, OH)},$ 3.64 (d, $J_{5,6}$ = 8.6, 5-H), 4.41 (d, $J_{6,5}$ = 8.3, 6-H), 5.25 (br s, 9-H), 6.20 (br t, $J_{14,13}$ = 4.5, 14-H). ¹³C NMR (75.5 MHz, 9-H), 6.20 (of t, $J_{14,13} = 4.3$, 14-H). C NMK (73.5 MHz, CDCl₃; contains ether): $\delta = \text{"+"21.62}$ and "+"27.36 [4-(CH₃)₂], "-"22.06 (C-12), "-"26.28 and "-"33.33 (C-11, C-13), "-"34.27 (because of relatively low intensity: C-4), "+"64.77 (C-6), "+"79.13 (C-5), "-"80.78, "-"87.27, "-"94.27 and "-"97.50 (C-2, C-3, C-7, C-8), "+"104.18 (C-12). 9), "-"120.99 (C-1), "+"139.79 (C-14), "-"145.03 (C-10). $C_{16}H_{18}O_2$ [M⁺]: calcd. 242.1306; the exact molecular mass (± 2 ppm; R = 10000) was confirmed by EI HRMS (70 eV).

5-Hvdroxv-4.4-dimethylbicvclo[8.4.0]tetradeca-1(14).7-diene-2,7-diyn-6-one (23). To a stirred solution of ButOMgBr [prepared by adding EtMgBr (2.0 M in THF, 4.35 mL, 8.70 mmol, 3.0 equiv.) to an ice-bath cooled solution of ButOH (0.83 mL, 8.7 mmol, 3.0 equiv.) in THF (15)] was added at -10 °C a solution of the diol 21 (702.1 mg, 2.90 mmol) in THF (12 mL). After 5 min, a solution of azodicarbonyldipiperidide (877.2 mg, 3.47 mmol, 1.2 equiv.) in THF (10 mL) was added. The reaction mixture was stirred at 0 °C for 30 min. After dilution with ether (20 mL), it was hydrolyzed by addition of brine (20 mL). After extraction with ether, the organic layer was dried over MgSO₄. The solvent was removed in vacuo at 0°C. Flash chromatography of the residue (etherpentane, 8:2) afforded 23 (380.6 mg, 55%). IR (CDCl₃): v = 3695 (very small), 3155, 2985, 2900, 2255, 2170, 1815, 1795, 1650, 1560, 1470, 1380, 1295, 1165, 1095 cm⁻¹. ¹H NMR (200 MHz, CDCl₃): $\delta = 1.09$ and 1.37 [2s, 4-(CH₃)₂], NMR (200 MHz, CDCl₃): $\delta = 1.09$ and 1.37 [2s, 4-(CH₃)₂], 1.67-1.84 (m, $12\cdot H_2$), 2.29 (td, $J_{13,12} = J_{13,14} = 5.53$, $13\cdot H_2$), 2.44 (td, $J_{11,12} = 6.4$, $^4J_{11,9} = 1.5$, $11\cdot H$), 3.84 (d, $J_{0H,5} = 4.9$, OH), 4.32 (d, $J_{5,OH} = 5.3$, $5\cdot H$), 5.47 (m_c, $9\cdot H$), 6.45 (td, $J_{14,13} = 4.7$, $^5J_{14,9} = 1.5$, $14\cdot H$). 13 C NMR (75.5 MHz, CDCl₃; contains ether): $\delta = "+"21.16$ and "+"26.47 [4-(CH₃)₂], "-"21.70 (C-12), "-"26.37 and "-"33.63 (C-11, C-13), "-"35.89 (because of relatively low intensity: C-4), "-"81.20, "-"94.16, "-"97.99 and "-"104.46 (C-2, C-3, C-7, C-8)" "+"82.98 (C-5)" +"102.48 (C-9)" "121.00 (C-1) C-7, C-8), "+" 82.98 (C-5), "+" 102.48 (C-9), "-" 121.00 (C-1), "+" 143.27 (C-14), "-" 153.39 (C-10), "-" 187.36 (C-6). $C_{16}H_{16}O_2$ [M⁺]: calcd. 240.1150; the exact molecular mass $(\pm 2 \text{ ppm}; R = 10000)$ was confirmed by EI HRMS (70 eV).

Z-5-[2-(3-Hydroxy-1-propynyl)-2-cyclohexenylidene]-2,2-dimethyl-3-pentyn-1-ol (24). To a solution of the bistriflate 17 (1.844 g, 4.72 mmol), $\operatorname{Cl_2Pd}(\operatorname{PPh_3})_2$ (166.4 mg, 2.41 mmol, 5 mol %) and CuI (90.1 mg, 0.472 mmol, 10 mol %) in ether (40 mL) was added at room temperature a solution of propargyl alcohol (291.3 mg, 5.19 mmol, 1.1 equiv.) in ether (11 mL) and then $\operatorname{Pr_2^iNH}$ (17 mL). The mixture was stirred for 3 h at this temperature. Then 2,2-dimethyl-3-butyn-1-ol (462.7 mg, 4.72

mmol, 1.0 equiv.) dissolved in Et₂O-Pr₂NH (3:1, 12 mL) was added. The resulting mixture was stirred at room temperature overnight and then hydrolyzed with brine (30 mL). After extraction with ether, the organic layer was dried over MgSO₄. The solvent was removed in vacuo at 0°C. The desired compound 24 was purified by flash chromatography (pentane-ether, 1:1). It could be liberated from some accompanying 'regioisomer' 19 by keeping the late fractions and rechromatographying several times the early fractions of the preceding separation; thus, we obtained absolutely isomerically pure 24 (446.2 mg, 39%). In a similar experiment, a single chromatography afforded 24 (629.9 mg, 62%) contaminated with 6% of the 'regioisomer' 19. IR (CDCl₃): v = 3500, 2970, 2930, 2870, 2255, 1450, 1425, 1395, 1365, 1285, 1200, 1045, 1020 cm⁻¹. ¹H NMR (200 MHz, CDCl₃): $\delta = 1.21$ [s, 2-CH₃)₂], 1.70–1.76 (m, 5'-H₂), 2.20–2.30 (m, 4'-H₂, 6'-H₂), 3.18 (s, OH), 3.40 (s, 1-H₂), 4.37 (s, 1"-H₂), 5.40 (br s, 5-H), 6.34 (poorly resolved td, $J_{3',4'} = 4.3$, ${}^5J_{3',5} = 1.1$, ${}^{3'}$ -H). 13 C NMR (125.7 MHz, CDCl₃): $\delta = "-"22.27$ (C-5'), "+"25.38 [2-(CH₃)₂], "-"26.85 and "-"33.13 (C-4', C-6'), "-"35.18 (because of relatively low intensity: C-2), "-"51.21 (C-3"), "-"71.57 (C-1), "-"79.87, "-"83.82, "-"88.38 and "-"100.95 (C-3, C-4, C-2", C-3"), "+"105.29 (C-5), "-"120.06 (C-2'), "-"142.09 (C-1'), "+"143.60 (C-3'). $C_{16}H_{20}O_2$ [M⁺]: calcd. 244.1463; the exact molecular mass $(\pm 2 \text{ ppm}; R = 10000)$ was confirmed by EI HRMS (70 eV).

Z-5-[2-(3-Oxo-1-propynyl)-2-cyclohexenylidene]-2,2-dimethyl-3-pentyn-1-al (25). To a stirred solution of the Dess-Martin periodinane (1.29 g, 3.04 mmol, 2.2 equiv.) in CH₂Cl₂ (20 mL) was added a solution of diol 24 (337.6 mg, 1.38 mmol) in CH₂Cl₂ (10 mL) at room temperature. After 30 min the reaction mixture was diluted with ether (20 mL) and poured into a saturated aqueous solution of NaHCO₃ containing a sevenfold excess of Na₂S₂O₃. The mixture was stirred for ca. 10 min until the solid was dissolved. Then the layers were separated. The organic layer was washed with water and dried over MgSO₄. The solvent was removed in vacuo at 0°C. Flash chromatography (pentane-ether, 85:15) of the residue afforded **25** (274.5 mg, 83%). IR (CDCl₃): v = 2980, 2935, 2865, 2810, 2255, 2190, 1730, 1655, 1570, 1455, 1420, 1390, 1250, 1195, 1135, 1070, 980, 895 cm⁻¹. ¹H NMR (200 MHz, $CDCl_3$): $\delta = 1.34$ [s, 2-CH₃)₂], 1.74–1.84 (m, 5'-H₂), 2.31–2.40 (m, 4'-H₂, 6'-H₂), 5.52 (hardly resolved d, ${}^{5}J_{5,3'} = 0.7$, 5-H), 6.70 (td, $J_{3',4'} = 4.5$, ${}^5J_{3',5} = 1.4$, 3'-H), 9.31 (s, 3"-H), 9.49 (s, 1-H). 13 C NMR (125.7 MHz, CDCl₃): $\delta = "-"21.76$ (C-5'), "+"22.44 [2-(CH₃)₂], "-"27.34 and "-"32.52 (C-4', C-6'), "-"43.48 (because of relatively low intensity: C-2), "-"81.78, "-"89.82, "-"94.63 and "-"97.08 (C-1", C-2", C-3, C-4), "+"106.29 (C-5), "-"118.87 (C-2'), "-"141.18 (C-1'), "+"150.10 (C-3'), "+"176.81 (C-3"), "+"197.99 (C-1). $C_{16}H_{16}O_2$ [M $^+$]: calcd. 240.1150; the exact molecular mass $(\pm 2 \text{ ppm}; R = 10000)$ was confirmed by EI HRMS (70 eV).

6,6-Dimethylbicyclo [8.4.0] tetradeca-1(14),9-diene-2,7-diyne-**4.5-diol (26):** (single diastereomer of unknown configuration). Cl₃Ti(DME)₂ was prepared by refluxing TiCl₃ (6.33 g, 41.0 mmol, 17.9 equiv.) in freshly distilled DME (160 mL) for 1.5 days. The Zn/Cu couple (8.118 g, 125.03 mmol, 54.6 equiv.) was added. The resulting mixture was refluxed for 5 h. At -45 °C a solution of the dialdehyde **25** (550.8 mg, 2.29 mmol) in DME (60 mL) was added by means of a syringe pump over 5 h. After 30 min, the reaction mixture was diluted with ether and hydrolyzed by adding a saturated aqueous solution of NaHCO₃ (30 mL). After filtration over celite, the aqueous layer was extracted with ether, the organic layer was washed with brine and dried over MgSO₄. The solvent was removed in vacuo at 0°C. Flash chromatography (pentane-ether, 45:55) of the residue gave **26** (443.9 mg, 80%). IR (CDCl₃): v = 3565, 2935, 2875, 1730, 1575, 1540, 1420, 1235, 1190, 1115, 1090 cm⁻¹. ¹H NMR (200 MHz, CDCl₃; contains ether): $\delta = 1.20$ and 1.29 [2s, 6-(CH₃)₂], 1.67 (tt, $J_{12,11} = J_{12,13} = 6.1$, 12-H₂), 2.20 (td, $J_{13,12} = J_{13,14} = 5.4$, 13-H₂), 2.29 (br t, $J_{11,12} = 6.2$, 11-H₂), 3.24 (br s, 2 × OH), 3.64 (d, $J_{4,5} = 8.7$, 4-H), 4.40 (d, $J_{5,4} = 8.6$, 5-H), 5.20 (br s, 9-H), 6.22 (t, $J_{14,13} = 4.4$, 14-H). ¹³C NMR (125.7 MHz, CDCl₃; contains ether): $\delta =$ "+"21.29 and "+"26.97 [6-(CH₃)₂], "-"21.94 (C-12), "-"26.24 and "-"33.05 (C-11, C-13), "-"34.49 (because of relatively low intensity: C-6), "+"64.20 (C-4), "+"79.16 (C-5), "-"81.26, "-"86.64, "-"89.75 and "-"102.66 (C-2, C-3, C-7, C-8), "+"105.82 (C-9), "-"120.52 (C-1), "+"139.98 (C-14), "-"142.33 (C-10). C₁₆H₁₈O₂ [M⁺]: calcd. 242.1306; the exact molecular mass (±2 ppm; R = 10000) was confirmed by EI HRMS (70 eV).

5-Hydroxy-6,6-dimethylbicyclo [8.4.0] tetradeca - 1(14),9-diene-2,7-diyn-4-one (28). To a stirred solution of Bu^tOMgBr [prepared by adding EtMgBr (2.0 M in THF, 4.0 mL, 8.0 mmol, 3.0 equiv.) to an ice-bath cooled solution of ButOH (0.77 mL, 8.0 mmol, 3.0 equiv.) in THF (15)] was added at -10 °C a solution of the diol **26** (650.4 mg, 2.68 mmol) in THF (15 mL). After 5 min, a solution of azodicarbonyldipiperidide (811.9 mg, 3.35 mmol, 1.2 equiv.) in THF (15 mL) was added. The reaction mixture was stirred at 0 °C for 30 min. After dilution with ether (30 mL) it was hydrolyzed by addition of brine (20 mL). After extraction with ether, the organic layer was dried over MgSO₄. The solvent was removed in vacuo at 0 °C. Flash chromatography of the residue (pentaneether, 8:2) afforded 28 (489.3 mg, 76%). IR (CDCl₃): v = 3470, 3155, 2975, 2935, 2870, 2340, 2255, 2195, 1655, 1600,1385, 1565, 1465, 1270, 1250 cm⁻¹. ¹H NMR (200 MHz, CDCl₃; contains ether and a little pentane): $\delta = 1.07$ and 1.34 [2s, 6-(CH₃)₂], 1.66–1.82 (m, 12-H₂), 2.32 (td, $J_{13,12}$ = $\overline{J}_{13,14} = 5.7$, 13-H₂), superimposed partly by 2.38 (td, $J_{11,12} = 5.8$, $^4J_{11,9} = 1.3$, 11-H₂), 3.81 (d, $J_{\text{OH},5} = 5.3$, OH), 4.31 (d, $J_{5,OH} = 5.3$, 5-H), 5.35 (only partly resolved td, $^4J_{9,11} = ^5J_{9,14} \approx 1.2$, 9-H), 6.59 (td, $J_{14,13} = 4.5$, $^5J_{14,9} = 1.5$, 14-H). 13 C NMR (125.7 MHz, CDCl₃; contains ether): $\delta = "+"21.05$ and "+"25.97 [6-(CH₃)₂], "-"21.36 (C-12), "-"26.89, "-"32.56 and "-"36.30 (C-6, C-11, C-13), "-"81.61, "-"89.77, "-"102.50 and "-"102.58 (C-2, C-3, C-7, C-8), "+" 83.57 (C-5), "+" 106.72 (C-9), "-" 119.52 (C-1), "-"141.65 (C-10), "+"147.48 (C-14), "-"187.85 (C-4). $C_{16}H_{16}O_2$ [M⁺]: calcd. 240.1150; the exact molecular mass $(\pm 2 \text{ ppm}; R = 10000)$ was confirmed by EI HRMS (70 eV)

Cycloaromatization/cyclization of dienediyne 23. Experiment A. At room temperature, NEt₃ (22 mL, 16 mg, 0.16 mmol, 1.0 equiv.), 1,4-cyclohexadiene (0.34 mL, 290 mg, 3.6 mmol, 23 equiv.) and methyl thioglycolate (30 µL, 35 mg, 0.33 mmol, 2.1 equiv.) were added under argon to a stirred solution of the hydroxyketone 23 (37.3 mg, 0.155 mmol) in C_6H_6 (1.5 mL). After 10 h, addition of brine (2 mL), extraction with ether, and drying over MgSO₄, the solvent was removed in vacuo. Flash chromatography (pentane-ether, 75:25) of the residue afforded in the early fractions 29/iso-29 as a 60:40 mixture of unassigned diastereomers (14.8 mg, 27%), in the middle fractions pure 30 (14.8 mg, 21%), and in the late fractions a 46:54 mixture (as determined from the 1-H integrals at $\delta_{in30} = 4.41$ and $\delta_{\text{in } iso-30} = 4.21$; 9.2 mg, 13%) of 30 and iso-30. The overall yield of 30 was therefore 19.0 mg (27%) and that of iso-30 was 4.9 mg (7%).

Experiment B. At room temperature, NEt₃ (20 μ L, 15 mg, 0.14 mmol, 1.0 equiv.) and methyl thioglycolate (27 μ L, 31 mg, 0.30 mmol, 2.1 equiv.) were added under argon to a stirred solution of the hydroxyketone **23** (33.9 mg, 0.141 mmol) in C₆H₆ (1.5 μ L). After 10 h, brine (2 μ L) was added. The same work-up procedure as in Experiment A delivered a 60:40 mixture of unassigned diastereomers **29** and iso-**29** (8.0 mg, 16%), pure **30** (12.6 mg, 20%) and a 16:84 mixture (12.5 mg,

19%) of **30** and *iso-***30**. Thus, we obtained a total of 14.7 mg (23%) of **30** and 10.4 mg (16%) of *iso-***30**.

From the combined 30/iso-30 mixtures of several cycloaromatizations like the ones described above we obtained—by enrichment in the late fractions of renewed attempts of purification by flash chromatography—a sufficiently pure sample of iso-30 for characterization.

Methyl - 2 - [(1,2,3,4,5,6,7,8 - octahydro - 7 - hydroxy - 8,8 - dimethyl-6-oxoanthracen-1-yl)thio | acetate: (60:40 mixture of unassigned diastereomers 29 and iso-29 as determined by integration of the 1'-H₂ resonances). IR (CDCl₃): v = 3500, 2935, 2870, 2255, 1720, 1605, 1500, 1435, 1385, 1365, 1275, 1190, 1160, 1130, 1085, 895 cm⁻¹. ¹H NMR (300 MHz, CDCl₃; slightly contaminated; for the numbering cf. Table 1): $\delta = 0.95$ and 1.58 {2s, 8-(CH₃)₂ [29], 0.96 and 1.57 {2s, 8-(CH₃)₂ [iso-**29**]}, 1.24–1.33 and 1.74–1.86 (2m, 3-H₂), 1.98–2.22 (m, 2-H₂, OH), 2.64–2.86 (m, 4-H₂), AB signal $\{\delta_A = 3.21, \delta_B = 3.35,$ $J_{AB} = 14.9$, 1'-H₂ [iso-**29**], AB signal $\{\delta_A = 3.23, \delta_B = 3.36,$ $J_{AB} = 15.1, 1'-H_2 [29]$, 3.74 (s, 5-H₂), 3.78 (s, CO₂Me), 4.17 $\{d, J_{7,OH} = 3.4, 7-H [iso-29]\}$, superimposed partly by 4.20 $\{d, J_{7,OH} = 3.4, 7-H [iso-29]\}$ $J_{7,OH} = 3.7, 7-H$ [29]}, 4.30 {poorly resolved t, $J_{1,2} = 3.8, 1-H$ [29*]}, superimposed partly by 4.31 {poorly resolved t, $J_{1,2} = 4.1$, 1-H [iso-29*]}, 6.86 (s, 10-H)**, 7.42 {s, 9-H** [iso-29]}, 7.43 {s, 9-H** [29]}; *,** assignments interchangeable. ¹³C NMR (75.5 MHz, CDCl₃; contains small peaks of contaminant(s) at $\delta = "-" 33.16$ and "+" 52.45; for the numbering cf. Table 1): $\delta = "-"18.62$ (C-3), "+"23.43 and "+" 24.19 {8-(CH₃)₂ [iso-29]}, "+" 23.62 and "+" 24.27 {8-(CH₃)₂ [29]}, "-" 28.29 and "-" 33.08* {C-2, C-4 [iso-29]}, "-" 28.34 and "-" 33.06* {C-2, C-4 [29]}, "-" 41.06, "-"41.09 and "-"42.19 (3 signals for 4 differents carbons, C-1', C-5), "-" 42.11 (because of relatively low intensity: C-8), "+"44.02 (CO₂CH₃), "+"52.37 {C-1 [**29**]}, "+"52.39 {C-1 [iso-29]}, "+"80.22 {C-7 [29]}, "+"80.32 {C-7 [iso-29]}, "+" 127.41 and "+" 129.45 $\{C-9, C-10 \text{ [iso-29]}\}$, "+" 127.43 and "+"129.41 {C-9, C-10 [29]}, "-"129.24 (double intensity), "-"134.85, "-"134.90, "-"136.48, "-"136.49, "-" 141.03 and "-" 141.10 (7 signals for 8 differents carbons: C-4a, C-8a, C-9a, C-10a), "-"171.11 {CO₂CH₃ [iso-29]}, "-"171.16 {CO₂CH₃ [29]}, "-"209.74 (C-6); * interchangeable. $C_{19}H_{24}O_4S$ [M⁺]: calcd. 348.1395; the exact molecular mass (± 2 ppm; R = 10000) was confirmed by EI HRMS (70 eV). C₁₉H₂₄O₄S (348.46); calcd.: C 65.49, H 6.94; found: C 65.74, H 7.18.

S',S'-{2,3,4,6,7,9-Hexahydro-7-hydroxy-6,6-dimethyl-8-oxo-¹H-benzo[a] azulene-4,5-diyl} bis[methyl-(2-sulfanylacetate)] (30): (single diastereomer of unknown configuration). IR $(CDCl_3)$: v = 3470, 2955, 2870, 2255, 1730, 1665, 1575, 1435, 1390, 1270, 1200, 1130, 1085, 1020 cm $^{-1}$. 1 H NMR (500 MHz, CDCl₃; slightly contaminated but not with a diastereomer; for the numbering—which exceptionally does not refer to the IUPAC name—cf. Table 1): $\delta = 0.79$ and 1.33 [2s, 8-CH₃)₂], 1.80–1.87 and *ca.* 2.24 – 2.32 (m, 3-H₂),^① $\stackrel{?}{=}$ 1.88–1.96 and 2.18 – ca. 2.24 (m, 2-H₂),^① $^{\circ}$ AB signal $[\delta_A = 2.79, \delta_B = 2.89,$ $J_{\rm AB}=17.9$, in addition split by $J_{\rm A,3-H(1)}{}^*=12.3$, $J_{\rm A,3-H(2)}{}^*=6.4$, $J_{\rm B,3-H(1)}{}^{**}=5.7$ (whereas $J_{\rm B,3-H(2)}{}^{**}\approx0$), $4{\rm -H_2}$], 0superimposes A part of AB signal ($\delta_{A} = 2.89$, $\delta_{B} = 3.22$, $J_{AB} = 17.8$, 5-H₂), AB signal ($\delta_{A} = 3.19$, $\delta_{B} = 3.36$, $J_{AB} = 14.5$, 1'-H₂), AB signal ($\delta_{A} = 3.66$, $\delta_{B} = 3.70$, $J_{AB} = 15.2$, 1"-H₂), 3.75 and 3.80 (2s, CO₂Me), 3.94 (d, $J_{OH,7} = 2.6$, OH), 4.18 (d, $J_{7,OH} = 2.5$, 7-H), 4.41 (br s, 1-H), 6.99 (s, 10-H); *, ** assignments interchangeable; $^{\odot}$ starting from δ_{1-H} (br s at 4.41 ppm) crosspeaks in a 500 MHz H,H-correlation spectrum allowed us to identify the chemical shifts of 2-H₂ (2 m at 1.88– 1.96 and 2.18-ca. 2.24) and, continuing from there, to recognize 3-H₂ (2 m at 1.80-1.87 and ca. 2.24-2.32) and ultimately 4-H₂ (AB signal, $\delta_A = 2.79$, $\delta_B = 2.89$); ²³ gem-relationship between these protons underlined by the occurrence of cross-

peaks in the 500 MHz one-bond C,H-correlation spectrum with pairwise the same ¹³C resonance; ³5-H₂ distinguished from 1'-H₂/1"-H₂ by means of a 500 MHz long-range C,Hcorrelation spectrum through the lack of a crosspeak with either of the CO₂CH₃ resonances in the first case and the occurrence of such crosspeaks in the second ($\delta_C = 170.72$) and third ($\delta_C = 169.96$) cases; ⁽⁴⁾distinguished because of the absence of a crosspeak in the 500 MHz one-bond C,H-correlation spectrum for $\delta_H = 3.94$ and the occurrence of such a crosspeak for $\delta_H=4.\overline{18}$ (with $\delta_C=80.21). \ ^{13}C$ NMR (APT spectrum, 125.7 MHz, CDCl₃; for the numbering—which exceptionally does not refer to the IUPAC name—cf. Table acceptionary does not refer to the ToTAC name—*tj.* Table 1): $\delta = "-"16.97 \text{ (C-3)}, ^{\odot} "+"18.42 \text{ and } "+"28.18 [8-(CH_3)_2], "-"27.20 (C-2), ^{\overline{0}} "-"30.30 (C-4), ^{\overline{0}} "-"33.10 and "-"34.42 (C-1', C-1"), "-"38.21 (C-8), ^{\overline{0}} "-"39.33 (C-5), ^{\overline{0}} "+"41.31 (C-1), ^{\overline{0}} "+"52.43 and "+"52.69 (2 \times CO_2CH_3), "+"80.21 (C-7), "+"124.00 (C-10), "-"125.29, "-"130.11, ""142.00 (C-10), "-"125.29, ""-"130.11, ""142.00 (C-10), "-"125.29, ""-"130.11, ""142.00 (C-10), "-"125.29, ""-"130.11, ""142.00 (C-10), "-"125.29, ""-"130.11, ""142.00 (C-10), ""-"125.29, ""-"130.11, ""142.00 (C-10), ""-"125.29, ""-"130.11, ""142.00 (C-10), ""-"125.29, ""-"125.29, ""-"125.29, ""-"125.29, ""-"125.29, ""-"125.29, ""-"125.29, ""-"125.29, ""-"125.29, ""-"125.29, ""-"125.29, ""-"125.29, ""-"125.29, ""-"125.29, ""-"125.29, ""-"125.$ "-"140.40, "-"143.69 and "-"144.28 (C-9, C-4a, C-8a, C-9a, C-10a), "-"169.96 and "-"170.72 ($2 \times COOCH_3$), "-"198.98 (C-6); ^① assigned because of the occurrence of crosspeaks in the 500 MHz one-bond C,H-correlation spectrum with the unambiguously assignable protons 1-H, 2-H₂, 3-H₂ and 4-H₂; ²distinguished through the absence of a crosspeak in the 500 MHz one-bond C,H-correlation spectrum with a proton in the case of $\delta_{\text{C}} = 38.21$ and the occurrence of such a crosspeak in the case of $\delta_C = 39.33$ (with $\delta_{\rm H} = 2.89$ and 3.22). $C_{22}H_{28}O_6S_2$ [M⁺]: calcd. 452.1327; the exact molecular mass (± 2 ppm; R = 10000) was confirmed by EI HRMS (70 eV). C₂₂H₂₈O₆S₂ (452.58); calcd.: C 58.38, H 6.24; found: C 58.51, H 6.42.

S',S'-{2,3,4,6,7,9-Hexahydro-7-hydroxy-6,6-dimethyl-8-oxo-¹H-benzo[a]azulene-4,5-diyl}bis[methyl-(2-sulfanylacetate)] (iso-30): (single diastereomer of unknown configuration). IR $(CDCl_3)$: v = 3500, 2955, 2870, 2255, 1730, 1665, 1600, 1575, 1545, 1435, 1390, 1280, 1200, 1135, 1080, 1020 cm⁻¹. ¹H NMR (500 MHz, CDCl₃; slightly contaminated; for the numbering—which exceptionally does not refer to the IUPAC name—cf. Table 1): $\delta = 0.81$ and 1.33 [2s, 8-CH₃)₂], 1.80–1.89 (!) and 2.16-2.22 (m, $2-H_2$), 0 1.80-1.89 (!) and 2.27-2.38 (m, 3-H₂),^① AB signal ($\delta_A = 2.69$, $\delta_B = 3.64$, $J_{AB} = 16.8$, 5-H₂),^② AB signal $[\delta_{A} = 2.79, \delta_{B} = 2.91, J_{AB} = 18.2, \text{ in addition split}$ by $J_{A,3-H(1)}^{*} = 12.0, J_{A,3-H(2)}^{*} = 6.6, J_{B,3-H(1)}^{*} = 6.4$ (whereas $J_{B,3-H(2)}^{*} * \approx 0$), $4 \cdot H_{2}$], AB signal $(\delta_{A} = 3.19, \delta_{B} = 3.36, J_{AB} = 14.7, 1' \cdot H_{2})$, AB signal $(\delta_{A} = 3.65, \delta_{B} = 3.71, J_{AB} = 4.77, 1' \cdot H_{2})$, AB signal $(\delta_{A} = 3.65, \delta_{B} = 3.71, J_{AB} = 4.77, 1' \cdot H_{2})$ 15.2, 1"-H₂), low-field branch of B part almost overlapped by 3.74 and 3.80 (2s, $2 \times \text{CO}_2\text{Me}$), 3.95 (d, $J_{\text{OH},7} = 2.6$, OH), $4.13 \text{ (d, } J_{7,OH} = 2.5, 7-H),$ $3 4.20 \text{ (m}_{c}, 1-H), 6.98 \text{ (s, } 10-H); *, **$ assignments interchangeable; $^{\odot}$ starting from δ_{1-H} (m_e at 4.20 ppm) crosspeaks in a 500 MHz H,H-correlation spectrum allowed us to identify the chemical shifts of 2-H₂ (2 m at 1.80– 1.89 and 2.16-2.22) and, continuing from there, to recognize $3-H_2$ (2 m at 1.80–1.89 and 2.27–2.38) and ultimately $4-H_2$ (AB signal, $\delta_A = 2.79$, $\delta_B = 2.91$); ²5-H₂ distinguished from 1'-H₂ and from 1"-H₂ by means of a 500 MHz long-range C,H-correlation spectrum through the lack of a crosspeak with either of the CO₂CH₃ resonances in the first case and the occurrence of such crosspeaks in the second ($\delta_{\rm C} = 169.94$) and third ($\delta_C = 170.64$) cases; ³assignment by analogy to compound 30. ¹³C NMR [APT spectrum, 125.7 MHz, CDCl₃; contains peaks of contaminant(s); for the numbering—which exceptionally does not refer to the IUPAC name-cf. Table 1]: $\delta = \text{``-''}16.96 \text{ (C-3)*, ``+''}17.61 \text{ and ``+''}28.10 [8-(CH₃)₂], ``-'''26.24 (C-2)*, ``-'''29.88 (C-4)*, ``-'''33.05 and ``-'''34.25 (C-1', C-1'')*, ``-'''38.49 and ``-'''38.94 (C-5, C-8)*,$ "+" 41.08 (C-1), "+" 52.42 and "+" 52.68 ($2 \times CO_2CH_3$), "+" 80.66 (C-7), "+" 123.78 (C-10), "-" 125.40, "-" 130.03, "-"140.44, "-"144.15 and "-"144.43 (C-9, C-4a, C-8a, C-9a, C-10a), (C-9), "-"169.94 and "-"170.64 (2 × COOCH₃), "-"199.06 (C-6); * assignments by analogy to

30 but tentative. $C_{22}H_{28}O_6S_2$ [M⁺]: calcd. 452.1327; the exact molecular mass (± 2 ppm; R = 10000) was confirmed by EI HRMS (70eV). No combustion analysis was performed due to lack of material.

Cycloaromatization of dienediyne 28. Experiment A. Two hours of a room temperature reaction under argon atmosphere between NEt₃ (22 µL, 16 mg, 0.16 mmol, 1.0 equiv.), 1,4-cyclohexadiene (0.34 mL, 290 mg, 3.6 mmol, 23 equiv.), methyl thioglycolate (31 µL, 36 mg, 0.34 mmol, 2.1 equiv.) and hydroxyketone **28** (38.2 mg, 0.159 mmol) in CH₂Cl₂ (2 mL) delivered—after work-up (as detailed for the cycloaromatization of hydroxyketone 23; flash chromatography with pentane-ether 7:3; vide supra)—34 (21.2 mg, 38%) in the early fractions and 35 in the late fractions (11.6 mg, 16%).

Experiment B. Similarly to Experiment A, NEt₃ (15 μ L, 11 mg, 0.11 mmol, 1.0 equiv.), methyl thioglycolate (0.20 mL, 230 mg, 2.2 mmol, 20 equiv.) and hydroxyketone 28 (26.6 mg, 0.111 mmol) in CH₂Cl₂ (1.3 mL) delivered after 10 h **34** (6.9 mg, 18%) and 35 (15.2 mg, 30%).

Methyl - [S - (1,2,3,4,5,6,7,8 - octahydro - 7 - hydroxy - 8,8 - dimethyl-6-oxophenanthren-4-yl)-(2-sulfanylacetate)] (34): (single diastereomer of unknown configuration). IR (CDCl₃): v =3485, 2935, 2870, 2260, 1720, 1600, 1480, 1440, 1410, 1385, 1365, 1275, 1190, 1125, 1080, 1010, 910 cm⁻¹. ¹H NMR (500 MHz, CDCl₃; for the numbering cf. Table 2): $\delta = 0.94$ and 1.56 [2s, 8-(CH₃)₂], 1.81–1.87 and 2.23–2.33 (2 m, 2-H₂), $^{\odot}$ 1.87–1.95 and 2.16–2.21 (2 m, 3-H₂), $^{\odot}$ AB signal ($\delta_{\rm A} = 2.77$, $\delta_{\rm B} = 2.87$, $J_{\rm AB} = 17.2$, in addition split by $J_{\rm A,2-H(1)} = 11.8^{*}$, $J_{\rm A,2-H(2)} = 6.0^{*}$, $J_{\rm B,2-H(1)} = 6.0^{**}$, whereas $J_{\rm B,2-H(2)} \approx 0^{**}$, $I_{\rm H_2}$, $\Omega_{\rm AB} = 14.4$, $I_{\rm H_2}$, $\Omega_{\rm AB} = 14.4$ AB signal ($\delta_A = 3.67$, $\delta_B = 4.29$, $J_{AB} = 22.1$, 5-H₂), 3.84 (d, $J_{\text{OH},7} = 4.0, \text{ OH},^{\odot} 3.86 \text{ (s, CO}_2\text{CH}_3), 4.12 \text{ (br s, 4-H)},^{\odot} 4.23$ (br d, $J_{7,OH} = 3.9$, 7-H),³ 7.07 (d, $J_{10,9} = 8.1$, 10-H)***, 7.30 (d, $J_{9,10} = 8.1$, 9-H)***; *, *** assignments interchangeable; ¹ starting from δ_{4-H} (br s at 4.12) crosspeaks in a 500 MHz H,H-correlation spectrum allowed us to identify the chemical shifts of 3-H₂ (2 m at 1.87-1.95 and 2.16-2.21) and, continuing from there, to recognize 2-H₂ (2 m at 1.81-1.87 and 2.23-2.33) and ultimately 1-H₂ (AB signal, $\delta_A = 2.77$, $\delta_{\rm B} = 2.87$); ²5-H₂ distinguished from 1'-H₂ by means of a 500 MHz long-range C,H-correlation spectrum through the occurrence of a crosspeak with the ketone resonance ($\delta_{\rm C}$ = 209.65) and not with the CO_2CH_3 resonance in the first case and with the CO_2CH_3 resonance ($\delta_C = 170.73$) and not with the ketone resonance in the second case; ³OH and 7-H distinguished through the occurrence of a crosspeak in the 500 MHz one-bond C,H-correlation spectrum between $\delta_{\rm C} = 79.73$ (C-7) and $\delta_{\rm H} = 4.23$ (7-H) and the absence of a crosspeak between $\delta_C = 79.73$ (C-7) and $\delta_H = 3.84$ (OH). ¹³C NMR (125.7 MHz gated decoupled; 75.5 MHz APT; CDCl₃; for the numbering cf. Table 2): $\delta = \text{``-''}17.37 \text{ (C-2),}^{\odot} \text{``+''}23.72 \text{ and }^{\circ} +^{\circ}24.63 \text{ [8-(CH₃)₂], $^{\circ}-^{\circ}27.38 \text{ (C-3),}^{\odot} \text{``-''}29.00 \text{ (C-1),}^{\odot} \text{``-''}33.24 \text{ (C-1),}^{\odot} \text{``-''}38.13 \text{ (C-5),}^{\odot} \text{``+''}42.39 \text{ (C-4),}^{\odot}$ "-"42.62 (C-8), "+"52.55 (CO_2CH_3) , "+"79.73 (C-7)"+" 124.74 and "+" 128.81 (C-9, C-10), "-" 129.97, "-" 132.90, "-" 136.54 and "-" 141.55 (C-4a, C-4b, C-8a, C-10a), "-"170.73 (CO₂CH₃), "-"209.65 (C-6); ^① assigned because of the occurence of crosspeaks in the 500 MHz onebond C,H-correlation spectrum with the unambiguously assignable protons $1-H_2$, $3-H_2$, 4-H, $5-H_2$, $1'-H_2$ and $1''-H_2$, no crosspeak for 2-H2 is detected; 2identified through the absence of a crosspeak in the 500 MHz one-bond C,H-correlation spectrum with any δ_H . $C_{19}H_{24}O_4S$ [M⁺]: calcd. 348.1395; the exact molecular mass (± 2 ppm; R = 10000) was confirmed by EI HRMS (70 eV). $C_{19}H_{24}O_4S$ (348.46); calcd.: C 65.49, H 6.94; found: C 65.34, H 7.20.

S',S'-(1,2,3,4,5,6,7,8-Octahydro-7-hydroxy-8,8-dimethyl-6oxophenanthrene-4,9-diyl) bis[methyl-(2-sulfanylacetate)] (35): (single diastereomer of unknown configuration). IR (CDCl₃): v = 3500, 2955, 2870, 2255, 1725, 1600, 1460, 1440, 1390, 1280,1195, 1130, 1090 cm⁻¹. ¹H NMR (500 MHz, CDCl₃; for the numbering cf. Table 2): $\delta = 0.94$ and 1.57 [2 s, 8-(CH₃)₂], 1.82–1.90 and 2.13–2.19 (2 m, 3- H_2), 1.90–1.95 and 2.25– 2.35 (2 m, 2-H₂),^① AB signal ($\delta_A = 2.56$, $\delta_B = 3.00$, $J_{AB} =$ 17.5, in addition split by $J_{A,2-H(1)} = 11.9^*$, $J_{A,2-H(2)} = 6.1^*$, $J_{B,2-H(1)} = 6.0^{**}$, whereas $J_{B,2-H(2)} \approx 0^{**}$, $1-H_2$), $^{\textcircled{\tiny }}$ AB signal $(\delta_A = 3.19, \delta_B = 3.34, J_{AB} = 14.4, 1'-H_2)$, $^{\textcircled{\tiny }}$ AB signal $(\delta_A = 3.65, \delta_B = 4.28, J_{AB} = 22.1, 5-H_2)$, low-field branch of A part superimposed entirely by the two central lines of extreme AB signal ($\delta_{\rm A} \approx \delta_{\rm B} \approx 3.67$, $J_{\rm AB}$ not determinable because the two outer lines are too small, 1"-H₂),² 3.75 and 3.86 (2 s, 2 \times CO₂CH₃), 3.83 (d, $J_{OH,7} = 4.0$, OH),³ 4.12 (br s, 4-H),¹ 4.21 (br d, $J_{7,OH} = 3.2$, 7-H),³ 7.36 (s, 10-H); *,*** assignments interchangeable, $^{\odot}$ starting from $\delta_{4\text{-H}}$ (br s at 4.12) crosspeaks in a 500 MHz H,H-correlation spectrum allowed us to identify the chemical shifts of 3-H₂ (2 m at 1.82-1.90 and 2.13-2.19) and, continuing from there, to recognize 2-H₂ (2 m at 1.90–1.95 and 2.25–2.35) and ultimately 1-H₂ (AB signal, $\delta_A=$ 2.56, $\delta_B=$ 3.00); $^{\odot}$ 5-H₂ distinguished from 1'-H₂/1"-H₂ by means of a 500 MHz long-range C,H-correlation spectrum through the lack of a crosspeak with either of the CO2CH3 resonances [and the presence of a crosspeak with $\delta_C = 209.33$ (C-6) in the first case and the occurrence of such crosspeaks in the second ($\delta_C = 170.67$) and third ($\delta_C = 170.03$) cases; ³OH and 7-H distinguished through the occurrence of a crosspeak in the 300 MHz one-bond C,H-correlation spectrum between $\delta_{\rm C} = 79.49$ (C-7) and $\delta_{\rm H} = 4.21$ (7-H) and the absence of such a crosspeak with $\delta_{\rm H} = 3.83$ (OH). ¹³C NMR (125.7 MHz, CDCl₃; for the numbering *cf.* Table 2): $\delta = \text{``-''}17.01 \text{ (C--2)},^{\text{(1)}}$ "+"23.66 and "+"24.45 [8-(CH₃)₂], "-"26.42 (C-3), $^{\odot}$ "-"26.70 (C-1), $^{\odot}$ "-"33.05 (C-1), $^{\odot}$ "-"34.86 (C-1"), $^{\odot}$ "-"37.95 (C-5), $^{\odot}$ "+"42.51 (C-4), $^{\odot}$ "-"42.87 (C-8), $^{\odot}$ " "+" 52.49 and "+" 52.66 (2 × CO_2CH_3), "+" 79.49 (C-7), "+" 124.06 (C-10), "-" 128.16, "-" 133.86, "-" 134.51, "-" 135.39 and "-" 142.09 (C-9, C-4a, C-8a, C-4b, C-10a), "-"170.03 and "-"170.67 ($2 \times CO_2CH_3$), "-"209.33 (C-6); ^① assigned because of the occurence of crosspeaks in the 300 MHz one-bond C,H-correlation spectrum with the unambiguously assignable protons 1-H2, 2-H2, 3-H2, 4-H, 5-H2, 1'-H₂, 1"-H₂; [©]identified through the absence of a crosspeak in the 300 MHz one-bond C,H-correlation spectrum. $C_{22}H_{28}O_6S_2$ [M⁺]: calcd. 452.1327; the exact molecular mass (± 2 ppm; R = 10000) was confirmed by EI HRMS (70 eV). C₂₂H₂₈O₆S₂ (452.58); calcd.: C 58.38, H 6.24; found: C 58.65, H 6.52.

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